# **One-step Library Preparation Method Supporting Skim-seq, Low Pass** Sequencing Approach for Genotype Imputation

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### Introduction

Recent advancements in next-generation sequencing (NGS) and bioinformatics have enabled whole-genome sequencing (WGS) to become a routine tool in both research and clinical applications. Low-coverage WGS, also known as skim-sequencing or "low-pass" sequencing, combined with imputation is a highly effective and cost-conscious alternative approach to microarray-based genotyping. Here, we demonstrate as a proof of concept, the use of Custom ExpressPlex<sup>TM</sup> 2.0, a one-step library preparation method, for skim-seq approach to genotype imputation on human samples. Custom ExpressPlex 2.0 was used to processed two individual human genomic DNA at three different total mass inputs in duplicate, generating a normalized 6-plex library pool. A 2 x 150 bp run on a NovaSeq X Plus sequenced the ExpressPlex libraries to  $\geq 20$  million paired-end reads per sample. Paired-end reads for each sample were aligned to the GRCh38 human reference genome, the precision and accuracy of SNP calls were determined for chromosome 1. The open-source GLIMPSE pipeline performed imputation using default settings. Our results show the utility of Custom ExpressPlex 2.0 for routine low-pass WGS applications, where we characterize multiplexing uniformity and genotype imputation accuracy on a collection of reference samples.

(30 minutes hands-on)

## — ExpressPlex Library Preparation Workflow









Figure 1. ExpressPlex 2.0 uses seqWell's high performance TnX<sup>™</sup> transposase that was specifically engineered for NGS library preparation. (A) ExpressPlex 2.0 library preparation kits tag input DNA with indexed adapters and amplify libraries all in a single reaction. Different full-length i7 indexed adapters tag the 96 DNA samples and barcoded libraries are amplified in separate wells, making for a highly efficient, one-step multiplexed library prep workflow. (B) Using the ExpressPlex 2.0 (96-well) kit, a 96-plex library can be prepared for library QC and sequencing in under 120 minutes, with less than 30 minutes of hands-on time. *Custom* ExpressPlex 2.0 used in the experiments and reported in this poster were performed using the same workflow as our standard **ExpressPlex 2.0 Library Preparation Kit.** 

### Methods

- Two individual human genomic DNA from the reference cohorts of CEPH/Utah and Han Chinese cohorts (Table 1) varying in mass input (10, 50, and 100 ng) were processed using Custom ExpressPlex 2.0 Library Preparation Kit.
- The 6-plex ExpressPlex library was sequenced on the NovaSeq X Plus. Sequencing data were individually down-sampled to 1M, 2M, 5M, 10M, and 20M random paired-end reads (0.1X, 0.2X, 0.5X, 1X, and 2X coverage, respectively) before variant calling and imputation.
- The open-source GLIMPSE pipeline (v2.0.0) performed imputation using default settings.

#### **Table 1.** Summary of hgDNA samples assessed in the study.

	Coriell ID	NIST ID	<b>Reference Cohort</b>	DNA Input Amount (ng)
			G001 CEPH/ Utah	10
	NA12878	HG001		50
				100
			Chinese	10
	NA24631	HG005		50
				100

### **Genotyping Imputation from Various Depth of Coverage**





Figure 2. The F1-score of low-pass WGS genotyping for two individual human genomic samples with Custom ExpressPlex 2.0 library prep on an Illumina NovaSeq X Plus System at various depth of coverage across three different DNA inputs.

The precision and accuracy of SNP calls were determined for chromosome 1. Prior analysis of a genetically diverse group of human samples sequenced at a low depth indicated that summary statistics of chromosome 1 statistically agreed with the imputation results from all autosomes. Figure 2 and Table 2 show the F1-score, recall, and precision of correct imputed genotype remains high despite various DNA input used at various coverage depth.

 
 Table 2. Summary of genotype imputation accuracy for
 chromosome 1 in HG001 and HG005 at various depth of coverage

Coriell ID (NIST ID)	Depth of Coverage	Recall	Precision	F1-score
	0.1X	0.84	0.80	0.82
NA12878 (HG001)	0.2X	0.88	0.83	0.86
	0.5X	0.93	0.84	0.88
	1 X	0.94	0.85	0.89
	2X	0.95	0.85	0.90
NA24631 (HG005)	0.1X	0.84	0.79	0.81
	0.2X	0.87	0.81	0.84
	0.5X	0.91	0.83	0.87
	1 X	0.93	0.84	0.88
	2X	0.94	0.84	0.89

### **Sequencing Metrics**

Table 3. Sequencing metrics summary of 6-plex Custom ExpressPlex 2.0 on the NovaSeq X Plus.

Avg PF Reads	Reads Aligned	Avg Mean Read Length	Avg Median Insert Size	Avg Duplication Rate	Mean Coverage Depth (X)
5.84 x 10 <sup>7</sup>	99.6%	140.3	216 nt	19.8%	2.78

### **Summary and Conclusions**

- Custom ExpressPlex 2.0 supports truly multiplexed and highly scalable construction of library pools for low-pass WGS through its one-step workflow (Figure 1).
- A high proportion of imputed calls (>98%) were identical to those in the truth data set (Figure 2, Table 2) despite various DNA input used at various coverage depth, confirming that high accuracy of genotyping can be achieved from a minimal sequencing data.





Figure 3. Imputation quality score for all genetic variants at different minor allele frequency (MAF) for chromosome 1 in the GIAB Consortium HG001 and HG005 at various depth of coverage across low (10 ng) and high (100 ng) DNA inputs. The imputation quality score is an estimate of imputation quality on a scale of 0 to 1, where 1 indicates that a genotype has been imputed with high certainty.

• Increasing depth of coverage from 0.1X to 1X significantly improves the imputation quality score for genetic variant MAF (Figure 3).

Custom ExpressPlex 2.0 is currently in Alpha Testing. For more information about this product or interest in joining the alpha program, please contact alpha@seqwell.com

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