Echo Acoustic Liquid Handling and Next Generation TnX™ Transposase-based Technologies Enable Miniaturized, Automated NGS Library Preparation for Ultra High-Throughput Sequencing

Sabina Gude¹, John Palys¹, John Fuller², Jack Leonard¹, Rebecca Feeley¹, Jenna Couture¹, Stella Huang¹

¹seqWell, Inc., Beverly, MA USA ²Beckman Coulter Life Sciences, Indianapolis, IN USA ²Beckman Coulter Life Sciences, Indianapolis, IN USA



Introduction

ExpressPlex™ 2.0 is the fastest library preparation chemistry available. Using seqWell's high performance TnX transposase - which was specially engineered for NGS library preparation - it performs auto-normalization of sample input, fragments input DNA into sizes suitable for Illumina sequencers, and tags the DNA with combinatorialdual-indexed adapters in a single step. Sequencing results in uniform read-count statistics for hundreds to thousands of samples. ExpressPlex 2.0 complements the Beckman Coulter Echo 525 by offering robust multiplexing capabilities for sequencing projects. It is designed to maximize throughput and data vield, as well as allows researchers to process thousands of simultaneously. This scalability is vital for large-scale efforts in synthetic biology, where the ability to sequence multiple samples in parallel can significantly accelerate discovery.

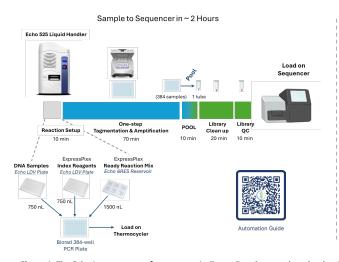
Here we demonstrate a proof-of-principle miniaturized (three-microliter) reaction setup on the Echo 525 for a single 384-well plate of pUC19 plasmid, resulting in highly consistent read counts (CV ~ 14% for 384/384 replicates). Using Echo's rapid contact-free transfers enabled by acoustic dispensing, the fast ExpressPlex 2.0 chemistry, and miniaturized reactions, it is possible to achieve economies of scale and ultra-high-throughput workflows in plasmid sequencing.

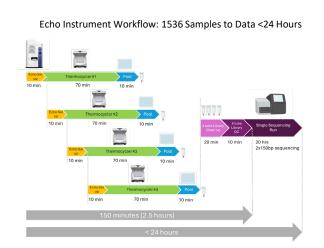
Summary

- Using Echo's rapid contact-free dispensing, the fast ExpressPlex 2.0 chemistry, and miniaturized reactions, it is possible to achieve economies of scale and ultra-highthroughput workflows in plasmid sequencing.
- The scale of miniaturization demonstrated is approximately one fifth of a standard ExpressPlex reaction, resulting in significant savings in reagent costs.
- This approach should also be valid for amplicon sequencing and other construct-validation applications in protein engineering and synthetic biology.

O segVell 2025. Figures created using BioRender.com. All Rights Reserved. Research use only. Not for use in diagnostics seelfwell, the segvelled logo, Expressible, and ITN are trademarks of segVelle, Inc. Beckman Coulter, the stylized logo, and the Beckman Coulter product and service marks mentioned herein are trademarks or registered trademarks of Beckman Coulter, line: in the United States and other countries. ECHO is a trademark or registered trademarks or Beckman Coulter, line: in the United States and other countries. The product of the Counter company. All other trademarks are the property of their respective owners. The method/application illustrated is for demonstration only, and is not validated by Beckman Coulter. Beckman Coulter makes no warranties of fitness for a particular purpose or merchantability or that the protocol is nonfiringing. All warranties are expressly disclaimed. Your use of the method is solely at your own risk, without recourse to Beckman Coulter. Not intended or validated for use in the diagnosis of disease or other conditions.

The Echo 525 Liquid Handler Enables Rapid Setup of Miniaturized ExpressPlex Reactions





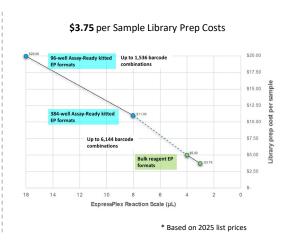


Figure 1: The Echo instrument performs acoustically-mediated contactless droplet dispensing. In the Echo/ExpressPlex 2.0 workflow, it rapidly and sequentially transfers DNA sample, Indexing Reagent, and Master Mix into a 384-well PCR plate. The plate is thermocycled, diluted in 10mM Tris, individual wells pooled into a single tube, purified by SPRI bead technology, quantitated via Agilent TapeStation, and sequenced on an Illumina MiSeq. pUC19 plasmid DNA (New England Biolabs) was diluted to 1.25 ng/uL and used in the Echo/ExpressPlex 2.0 library prep workflow.

Data and Analysis

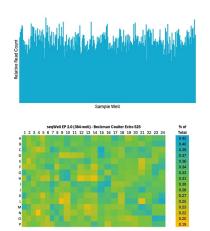


Figure 2: The CV of read counts across all wells is < 14% and there are no failed wells.

Figure 3: A color gradient image of read counts is even and devoid of systematic anomalies.

100% of Plasmids were Assembled & Circularized

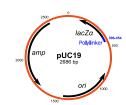


Figure 4: Bioinformatics analysis was carried out on seqWell's SNAP pipeline. When downsampled to 2,000 read pairs per sample, all plasmids were successfully assembled *de novo*.

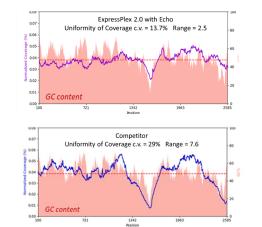


Figure 5.: The benefits of the ExpressPlex 2.0 engineered transposase are seen in reduced insertion bias and greater uniformity of coverage. The figure on the top depicts a typical outcome for the Echo-derived 3 uL sequencing libraries. GC content of the pUC19 genome is shown in red, with normalized coverage overplotted. The figure on the bottom depicts the same parameters for libraries derived using a competitor kit under standard conditions. (c.v. = coefficient of variation; Range = average of the three highest read counts divided by the average of the lowest three read counts.)